WEST Search History

Hide Items | Restore | Clear | Cancel

DATE: Monday, June 28, 2004

Hide?	Set Name	Query	Hit Count
		DB=USPT; PLUR=YES; OP=AND	
☐	L1	(hyperimmune or hyper-immune or hyper-immun$ or hyperimmun$) and (anthrax or anthrac$)	99
☐	L2	(hyperimmune or hyper-immune or hyper-immun$ or hyperimmun$) same (anthrax or anthrac$)	0
☐	L3	(hyperimmune or hyper-immune or hyper-immun$ or hyperimmun$).ti,ab,clm. and (anthrax or anthrac$)	9
☐	L4	(hyperimmune or hyper-immune or hyper-immun$ or hyperimmun$).ti,ab,clm. and (anthrax or anthraces$ or anthracsis$)	0
☐	L5	(hyperimmune or hyper-immune or hyper-immun$ or hyperimmun$) and (anthrax or anthraces$ or anthracsis$)	14
☐	L6	(hyperimmune or hyper-immune or hyper-immun$ or hyperimmun$) and (anthrax or anthraces$ or anthracsis$).clm.	2
☐	L7	(anthrax or anthraces$ or anthracsis$).clm.	30
		DB=PGPB,USPT,USOC,EPAB,JPAB,DWPI,TDBD; PLUR=YES; OP=AND	
☐	L8	antipa or anti-pa or antipa63 or anti-pa63 or antipa$3 or anti-pa$3	2762
☐	L9	L8 and anthrax	36

END OF SEARCH HISTORY

passive hyperimmune therapy

WEST Search History

Hide Items | Restore | Clear | Cancel

DATE: Monday, June 28, 2004

Hide?	Set Name	Query	Hit Count
		DB=PGPB,USPT,USOC,EPAB,JPAB,DWPI,TDBD; PLUR=YES; OP=AND	
☐	L1	(antianthrax or anti-anthrax).ti,ab,clm.	10
		DB=EPAB; PLUR=YES; OP=AND	
☐	L2	RU-2161985-C1.did.	0
☐	L3	AU-9932191-A.did.	0
☐	L4	WO-9950439-A2.did.	1
		DB=PGPB,USPT,USOC,EPAB,JPAB,DWPI,TDBD; PLUR=YES; OP=AND	
☐	L5	(antianthrax or anti-anthrax) not l1	24
		DB=EPAB; PLUR=YES; OP=AND	
☐	L6	RU-2161985-C1.did.	0
		DB=PGPB,USPT,USOC,EPAB,JPAB,DWPI,TDBD; PLUR=YES; OP=AND	
☐	L7	(anthrax or anthrac$).ti,ab,clm.	14288
☐	L8	L7 not l1 or l5	14284
☐	L9	L7 not l1 not l5	14260
☐	L10	L9 same (ivig or igg or gamma or gammaglobulin or ig or immune or immunoglobulin or antibodies or immunotherapy or scab or scfv or antiserum or antisera or anti-sera or anti-serum or polyclonal or poly-clonal).ti,ab,clm.	499
☐	L11	L10 same (method or process).ti,ab,clm.	138
☐	L12	(anthrax$ or $anthrax) not l1 not l4 not l5 not l10 not l11	2358
☐	L13	L12 same (ivig or igg or gamma or gammaglobulin or ig or immune or immunoglobulin or antibodies or immunotherapy or scab or scfv or antiserum or antisera or anti-sera or anti-serum or polyclonal or poly-clonal)	399
☐	L14	L12 near10 (ivig or igg or gamma or gammaglobulin or ig or immune or immunoglobulin or antibodies or immunotherapy or scab or scfv or antiserum or antisera or anti-sera or anti-serum or polyclonal or poly-clonal)	64

END OF SEARCH HISTORY

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Search Results - Record(s) 1 through 10 of 10 returned.

☐ 1. WO003063768A2. 25 Oct 02. 07 Aug 03. COMPOSITIONS AND METHODS DIRECTED TO ANTHRAX TOXIN. SOLTIS, DANIEL A, et al. A61K00/;.

☐ 2. RU 2214836C. Dry equine anti-anthrax globulin. BOGACHEVA, V V, et al. A61K039/07 A61K039/40.

☐ 3. RU 2214834C. Treating generalized form of anthrax infection. AMOSOV M YU,, et al. A61K039/07 A61K039/40 A61P031/00.

☐ 4. RU 2161985C. Preparation of semiproduct of equine anti-anthrax globulin. KOMISSAROV, A V, et al. A61K039/40.

☐ 5. RU 2142009C. Method of maintenance of anthrax vaccine strain sti-1. AMOSOV M YU,, et al. A61K039/07 C12N001/20 C12N001/20 C12R001:07.

☐ 6. WO 9950439A. Screening for modulators and mimetics of the lethal factor of anthrax toxin, useful as anticancer and anti-anthrax agents. DUESBURY, N, et al. C07H021/04 C12N009/12 C12Q001/00 C12Q001/37 C12Q001/48 G01N033/48 G01N033/50 G01N033/53 G01N033/567 G01N033/574 G06F019/00.

☐ 7. RU 1347224C. Prodn. of anti-anthrax serum - by two=stage hyper-immunisation with introduction of antigen five times subcutaneously in first stage. MANICHEV, A A, et al. A61K039/40.

☐ 8. SU 1807079A. Prodn of atoxigenic variants of anthrax stimulant - by inoculation of spores into nutrient medium incubating with elimination of toxigenicity plasmids, seeding and screening. LIKHOLETOV, S M, et al. C12N001/20.

☐ 9. SU 1791449A. Bacillus anthracis strain cultivation - by aerating aq. medium contg. beef acidic hydrolysate and salts at specific rate for 27-29 h, then leaving for 19-21 h.. BAKULOV, I A, et al. A61K039/10 C12N003/00.

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Terms	Documents
(antianthrax or anti-anthrax).ti,ab,clm.	10

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Search Results - Record(s) 1 through 24 of 24 returned.

☐ 1. 20040116370. 26 Aug 03. 17 Jun 04. Retroductal salivary gland genetic vaccination. Tucker, Sean, et al. 514/44; 424/93.2 514/150 A61K048/00 A61K031/655.

☐ 2. 20040096821. 29 Dec 03. 20 May 04. Detection of micro-organisms. Keenan, Elizabeth Ann, et al. 435/5; 435/7.22 435/7.31 435/7.32 C12Q001/70 G01N033/53 G01N033/569 G01N033/554.

☐ 3. 20040072241. 08 Jan 03. 15 Apr 04. Methods for detecting B. anthracis infection. Valkirs, Gunars Edwin, et al. 435/7.1; 422/61 435/975 G01N033/53.

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☐ 5. 20040023897. 12 Nov 02. 05 Feb 04. Methods for preventing or treating disease mediated by toxin-secreting bacteria. Caplan, Michael J.. 514/29; 514/153 514/192 514/200 514/253.08 514/313 A61K031/7048 A61K031/65 A61K031/43 A61K031/545 A61K031/496 A61K031/47.

☐ 6. 20040023266. 11 Mar 03. 05 Feb 04. Methods and compositions for aptamers against anthrax. Vivekananda, Jeevalatha, et al. 435/6; 435/252.31 C12Q001/68 C12N001/20.

☐ 7. 20040009945. 21 May 03. 15 Jan 04. Anthrax vaccine. Lee, John S., et al. 514/44; 435/320.1 536/23.7 A61K048/00 C12N015/74 C07H021/04.

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☐ 14. 20030054423. 19 Sep 01. 20 Mar 03. Attachment of biomolecules to hydrophobic surfaces. Anderson, George P., et al. 435/7.92; 436/518 G01N033/53 G01N033/537 G01N033/543.

☐ 15. 20020082386. 30 Apr 01. 27 Jun 02. Anthrax specific antibodies. Mangold, Beverly Lynn, et

al. 530/350; 424/184.1 A61K039/00 A61K039/38 C07K001/00 C07K014/00 C07K017/00.

☐ 16. 6713272. 19 Sep 01; 30 Mar 04. Attachment of biomolecules to hydrophobic surfaces. Anderson; George P., et al. 435/7.92; 435/69.7 435/7.94 436/518 436/523 436/524 530/387.3. G01N033/53.

☐ 17. 6569630. 15 Oct 01; 27 May 03. Methods and compositions for aptamers against anthrax. Vivekananda; Jeevalatha, et al. 435/6; 435/91.2 536/22.1 536/23.1. C12Q001/68 C12P019/34 C07H021/04 C07H021/02.

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☐ 20. 6303316. 30 Jun 00; 16 Oct 01. Organic semiconductor recognition complex and system. Kiel; Johnathan L., et al. 435/6; 435/7.1 435/91.2 436/94 536/23.1. C12Q001/68 C12P019/34 C07H021/04.

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Terms	Documents
(antianthrax or anti-anthrax) not L1	24

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Search Results - Record(s) 1 through 30 of 30 returned.

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☐ 4. 6716823. 23 Mar 00; 06 Apr 04. Noninvasive genetic immunization, expression products therefrom, and uses thereof. Tang; De-chu C., et al. 514/44; 424/93.21 435/320.1 435/375. A61K031/70 A61K048/00 C12N015/00 C12N005/00.

☐ 5. 6679419. 01 Feb 02; 20 Jan 04. Mailbox. Sarracino; Maximo. 232/17; 422/186.3 422/24. B65D091/00.

☐ 6. 6592872. 15 Sep 97; 15 Jul 03. Targeting antigens to the MHC class I processing pathway with an anthrax toxin fusion protein. Klimpel; Kurt, et al. 424/197.11; 424/183.1 424/184.1 424/192.1 424/193.1 424/195.11 424/236.1 424/246.1 514/2 514/885 530/323 530/350 530/402 530/403 530/825. A61K039/07 A61K039/02 C07K014/32 C07K019/00.

☐ 7. 6588650. 13 Nov 01; 08 Jul 03. Anthrax detecting envelope system. Polidori; Anthony. 229/71; 229/120.32 229/72. B65D027/04.

☐ 8. 6573245. 28 Apr 99; 03 Jun 03. Modified polysaccharide adjuvant-protein antigen conjugates, the preparation thereof and the use thereof. Marciani; Dante J.. 514/25; 424/185.1 424/193.1 424/194.1 514/42 514/54 514/55 514/61. A61K031/70 A61K039/00.

☐ 9. 6569630. 15 Oct 01; 27 May 03. Methods and compositions for aptamers against anthrax. Vivekananda; Jeevalatha, et al. 435/6; 435/91.2 536/22.1 536/23.1. C12Q001/68 C12P019/34 C07H021/04 C07H021/02.

☐ 10. 6541001. 21 Aug 00; 01 Apr 03. Vaccine composition and method of using the same. Gallili; Gilad, et al. 424/184.1; 424/204.1 424/214.1 435/5. A61K039/00 A61K039/12 A61K039/17 C12Q001/70.

☐ 11. 6524846. 05 Nov 01; 25 Feb 03. Biological toxin detection system for mailed materials. Robinson, Jr.; William L.. 435/287.4; 422/86 435/287.7 435/288.7 436/111. C12M001/34.

☐ 12. 6503906. 21 Feb 02; 07 Jan 03. Method for optimizing ciprofloxacin treatment of anthrax-exposed patients according to the patient's characteristics. Lee; Ren-Jin. 514/253.05; 514/253.07

514/253.08 514/885. A61K031/495.

☐ 13. 6488900. 19 Oct 99; 03 Dec 02. Method and apparatus for air purification. Call; Charles J., et al. 422/173; 422/120 422/122 422/125 422/174 422/177 422/180. B01D053/34 A62B011/00.

☐ 14. 6488872. 21 Jul 00; 03 Dec 02. Microfabricated devices and method of manufacturing the same. Beebe; David J., et al. 264/31; 264/139 264/240 264/254 264/255 264/267 264/34 264/35 264/459 264/463 422/50 422/68.1. E04B001/16.

☐ 15. 6440405. 06 Jun 00; 27 Aug 02. Quaternary ammonium functionalized dendrimers and methods of use therefor. Cooper; Stuart L., et al. 424/78.17; 424/486 424/719 424/DIG.16 564/281. A61K031/74 A61K047/48 A61K033/02 C07C211/62.

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☐ 17. 6424864. 19 May 00; 23 Jul 02. Method and apparatus for wave therapy. Matsuura; Masayuki. 607/3; 607/66 607/76. A61N001/32.

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☐ 19. 6316197. 01 Feb 00; 13 Nov 01. Method of diagnosing of exposure to toxic agents by measuring distinct pattern in the levels of expression of specific genes. Das; Rina, et al. 435/6; 435/91.2 435/91.5 536/24.31. C12Q001/68 C12P019/34 C07H021/04.

☐ 20. 6309633. 19 Jun 99; 30 Oct 01. Amphiphilic drug-oligomer conjugates with hydroyzable lipophile components and methods for making and using the same. Ekwuribe; Nnochiri, et al. 424/85.1; 424/193.1 424/194.1 424/85.2 424/85.4 424/94.3 435/188 514/12 514/2 514/21 514/3 514/476 514/506 514/579 514/613 514/715 514/8 530/303 530/345 530/405 530/406 530/409 530/410 530/411. A61K009/107 A61K038/17 A61K038/28 A61K039/385 C07K001/113.

☐ 21. 6303316. 30 Jun 00; 16 Oct 01. Organic semiconductor recognition complex and system. Kiel; Johnathan L., et al. 435/6; 435/7.1 435/91.2 436/94 536/23.1. C12Q001/68 C12P019/34 C07H021/04.

☐ 22. 6022855. 14 Sep 95; 08 Feb 00. Methods and reagents for inhibiting furin endoprotease. Thomas; Gary, et al. 514/12; 530/330 530/350. C07K014/81 C07K004/12 C07K005/10 A61K038/55.

☐ 23. 5874088. 05 Jan 95; 23 Feb 99. Deletion mutants of cholera vaccines expressing heterologous antigens. Mekalanos; John J.. 424/200.1; 424/203.1 424/235.1 424/261.1 435/243 435/252.1 435/252.3 435/69.3 435/909. A61K039/106 C12N001/21.

☐ 24. 5747028. 07 Jun 95; 05 May 98. Immunizing compositions comprising Vibrio cholerae expressing heterologous antigens. Calderwood; Stephen B., et al. 424/93.2; 435/252.3 435/69.1 435/69.3. C12N001/21 A61K039/106.

☐ 25. 5728385. 12 Aug 93; 17 Mar 98. Method and composition for an early vaccine to protect against both common infectious diseases and chronic immune mediated disorders or their sequelae. Classen; John Barthelow. 424/201.1; 424/184.1 424/202.1 424/203.1 424/212.1 424/217.1 424/218.1

424/219.1 424/224.1 424/227.1 424/228.1 424/230.1 424/233.1 424/234.1 424/244.1 424/245.1 424/246.1 424/247.1 424/249.1 424/254.1 424/258.1 424/261.1. A61K039/02 A61K039/12 A61K039/116 A61K039/295.

☐ 26. 5723283. 31 May 95; 03 Mar 98. Method and composition for an early vaccine to protect against both common infectious diseases and chronic immune mediated disorders or their sequelae. Classen; John Barthelow. 435/4; 424/184.1 424/204.1 424/234.1. C12Q001/02 A61K039/00 A61K039/02 A61K039/12.

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☐ 28. 5677274. 25 Jun 93; 14 Oct 97. Anthrax toxin fusion proteins and related methods. Leppla; Stephen H., et al. 514/2;. A61K039/00.

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☐ 30. 4781041. 27 Oct 86; 01 Nov 88. Apparatus for cleaning garments and soft goods contaminated with nuclear, chemical and/or biological contaminants. Fowler; David E.. 68/18F; 68/18R 976/DIG.376. D06F043/08.

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Terms	Documents
(anthrax or anthraces$ or anthracsis$).clm.	30

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First Hit

L1: Entry 7 of 10 File: DWPI Nov 20, 1995

DERWENT-ACC-NO: 1996-275528
DERWENT-WEEK: 199628

TITLE: Prodn. of anti-anthrax serum - by two=stage hyper-immunisation with introduction of antigen five times subcutaneously in first stage

INVENTOR: MANICHEV, A A; ROMANOV, G I ; SALENKO, L S

PATENT-ASSIGNEE: VETERINARY PREPARATS RES INST (VETER)

PRIORITY-DATA: 1986SU-4066843 (May 8, 1986)

Search Selected | Search ALL | Clear

PATENT-FAMILY:

	PUB-NO	PUB-DATE	LANGUAGE	PAGES	MAIN-IPC
☐	RU 1347224 C	November 20, 1995		005	A61K039/40

APPLICATION-DATA:

PUB-NO	APPL-DATE	APPL-NO	DESCRIPTOR
RU 1347224C	May 8, 1986	1986SU-4066843	

INT-CL (IPC): A61 K 39/40

ABSTRACTED-PUB-NO: RU 1347224C
BASIC-ABSTRACT:

Prodn. of an anti-anthrax serum comprises hyperimmunisation of antigen producers with anthrax strain STI-1 and subsequent isolation of the desired prod. Hyperimmunisation is carried out in 2 stages. In the first stage the antigen is introduced 5 times, s.c. and intradermally, and in the second stage 8 times s.c. with doses increasing from 8.0-12.0 to 78.0-126.0 multiplied by 109 microbe cells, where the interval between injections is 3-4 days. In the first stage, the s.c. injection doses are increased from 0.4-0.6 to 4.0-6.0 multiplied by 109 microbe cells, and the intradermal dose is 0.4-0.6 multiplied by 109 microbe cells.

USE - The method is useful in veterinary practice.

ADVANTAGE - The serum has increased activity and specificity, and is obtd. more quickly (43-57 days as opposed to 59-109 days) and easily than in prior art.

ABSTRACTED-PUB-NO: RU 1347224C
EQUIVALENT-ABSTRACTS:

CHOSEN-DRAWING: Dwg.0/0

DERWENT-CLASS: B04 C06 D16
CPI-CODES: B04-B04C1; C04-B04C1; B04-F10; C04-F10; B14-S11B; C14-S11B; B14-S12; C14-C14-S12; D05-H07;

First Hit

L1: Entry 8 of 10 File: DWPI Apr 7, 1993

DERWENT-ACC-NO: 1994-198606
DERWENT-WEEK: 199424

TITLE: Prodn of atoxigenic variants of anthrax stimulant - by inoculation of spores into nutrient medium incubating with elimination of toxigenicity plasmids, seeding and screening

INVENTOR: LIKHOLETOV, S M; LIPNITSKII, A V ; SHELOKHOVICH, A I

PATENT-ASSIGNEE: VOLG ANTI-PLAGUE RES INST (VLANR)

PRIORITY-DATA: 1990SU-4791684 (February 14, 1990)

Search Selected | Search ALL | Clear

PATENT-FAMILY:

	PUB-NO	PUB-DATE	LANGUAGE	PAGES	MAIN-IPC
☐	SU 1807079 A1	April 7, 1993		004	C12N001/20

APPLICATION-DATA:

PUB-NO	APPL-DATE	APPL-NO	DESCRIPTOR
SU 1807079A1	February 14, 1990	1990SU-4791684	

INT-CL (IPC): C12N 1/20

ABSTRACTED-PUB-NO: SU 1807079A
BASIC-ABSTRACT:

Method involves inoculation of spores into a nutrient medium, incubation under conditions which ensure elimination of toxigenicity plasmids, with subsequent seeding and screening of the colonies in a dense nutrient medium. The dense nutrient nutrient medium for seeding includes 15-25% therapeutic anti-anthrax serum. The cell cell concn. in seeding with spores is 10xpower 5-10xpower 3, and incubation is carried out at 37 deg.C in an atmos. of 6-25% CO2 over 18-24 hr.

The test culture is pref. inoculated into a semi-synthetic medium of known compsn. (Haines et al., 1965) with addn. of agarose. The medium ensures production of isolated colonies of atoxic variants of B. anthracis. The seeds are incubated at 27 deg.C for 1824 hr. in an atmos. of 6-25% COs. The seed material is applied to dense medium in a dose not less than 10xpower 5-10xpower 6 cells.

USE/ADVANTAGE - Used in medicine, i.e. in microbiology and immunology. The method is quicker and simpler. The method provides a means of obtaining atoxigenic variants of B. anthracis, which are useful for genetic studies (in studying signs of toxin formation in microbes), for introduction into plasmid-free strains of any other plasmids with useful properties, and in studying the ratio of the number of Tox super + (immunogenic) and Tox super - (non-immunogenic) cells in B. anthracis

culture populations for use as vaccines.

ABSTRACTED-PUB-NO: SU 1807079A
EQUIVALENT-ABSTRACTS:

CHOSEN-DRAWING: Dwg.0/0

DERWENT-CLASS: B04 D16
CPI-CODES: B04-F10; D05-A04; D05-H04; D05-H07; D05-H09;

First Hit

L1: Entry 8 of 10 File: DWPI Apr 7, 1993

DERWENT-ACC-NO: 1994-198606
DERWENT-WEEK: 199424

TITLE: Prodn of atoxigenic variants of anthrax stimulant - by inoculation of spores into nutrient medium incubating with elimination of toxigenicity plasmids, seeding and screening

INVENTOR: LIKHOLETOV, S M; LIPNITSKII, A V ; SHELOKHOVICH, A I

PATENT-ASSIGNEE: VOLG ANTI-PLAGUE RES INST (VLANR)

PRIORITY-DATA: 1990SU-4791684 (February 14, 1990)

Search Selected | Search ALL | Clear

PATENT-FAMILY:

	PUB-NO	PUB-DATE	LANGUAGE	PAGES	MAIN-IPC
☐	SU 1807079 A1	April 7, 1993		004	C12N001/20

APPLICATION-DATA:

PUB-NO	APPL-DATE	APPL-NO	DESCRIPTOR
SU 1807079A1	February 14, 1990	1990SU-4791684	

INT-CL (IPC): C12N 1/20

ABSTRACTED-PUB-NO: SU 1807079A
BASIC-ABSTRACT:

Method involves inoculation of spores into a nutrient medium, incubation under conditions which ensure elimination of toxigenicity plasmids, with subsequent seeding and screening of the colonies in a dense nutrient medium. The dense nutrient nutrient medium for seeding includes 15-25% therapeutic anti-anthrax serum. The cell cell concn. in seeding with spores is 10xpower 5-10xpower 3, and incubation is carried out at 37 deg.C in an atmos. of 6-25% CO2 over 18-24 hr.

The test culture is pref. inoculated into a semi-synthetic medium of known compsn. (Haines et al., 1965) with addn. of agarose. The medium ensures production of isolated colonies of atoxic variants of B. anthracis. The seeds are incubated at 27 deg.C for 1824 hr. in an atmos. of 6-25% COs. The seed material is applied to dense medium in a dose not less than 10xpower 5-10xpower 6 cells.

USE/ADVANTAGE - Used in medicine, i.e. in microbiology and immunology. The method is quicker and simpler. The method provides a means of obtaining atoxigenic variants of B. anthracis, which are useful for genetic studies (in studying signs of toxin formation in microbes), for introduction into plasmid-free strains of any other plasmids with useful properties, and in studying the ratio of the number of Tox super + (immunogenic) and Tox super - (non-immunogenic) cells in B. anthracis

culture populations for use as vaccines.

ABSTRACTED-PUB-NO: SU 1807079A
EQUIVALENT-ABSTRACTS:

CHOSEN-DRAWING: Dwg.0/0

DERWENT-CLASS: B04 D16
CPI-CODES: B04-F10; D05-A04; D05-H04; D05-H07; D05-H09;

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L1: Entry 9 of 10 File: DWPI Jan 30, 1993

DERWENT-ACC-NO: 1994-125400
DERWENT-WEEK: 199415

TITLE: Bacillus anthracis strain cultivation - by aerating aq. medium contg. beef acidic hydrolysate and salts at specific rate for 27-29 h, then leaving for 19-21 h. h.

INVENTOR: BAKULOV, I A; CHISLOV YU, V ; GAVRILOV, V A

PATENT-ASSIGNEE: GOSAGROPROM RES INST (GOSAR)

PRIORITY-DATA: 1988SU-3199979 (May 23, 1988)

Search Selected Search ALL Clear

PATENT-FAMILY:

	PUB-NO	PUB-DATE	LANGUAGE	PAGES	MAIN-IPC
☐	SU 1791449 A1	January 30, 1993		004	C12N003/00

APPLICATION-DATA:

PUB-NO	APPL-DATE	APPL-NO	DESCRIPTOR
SU 1791449A1	May 23, 1988	1988SU-3199979	

INT-CL (IPC): A61K 39/10; C12N 3/00

ABSTRACTED-PUB-NO: SU 1791449A
BASIC-ABSTRACT:

Process time needed to obtain spore material for anthrax vaccine prodn. can be cut to at least one-third.

A Bacillus anthracis strain developed at the national vaccine research institute (No. VNIIVVM) is grown on an aq. nutrient medium of the following compsn.,: beef acidic hydrolysate (60-80 ml); (NH4)2SO4 (1.5-2.5 g); disubstituted potassium phosphate (3.0-12.0 g); MgSO4 (0.15-0.25 g); propyleneglycol (0.08-0.12 ml); sodium citrate (0.8-1.2 g); distilled water (balance to 1 l). Cultivation proceeds for 46-50 h. For the first 27-29 h the culture is aerated by bubbling through air without mechanical stirring at an aeration rate of 3.5+-0.2 mmoles of dissolved O2 per 1 per per h.

Large amts. of spore material for anti-anthrax vaccine prodn. can be obtd. using the the technique, which reduces labour costs through mechanisation of the process. As a result spore material prodn. time is cut by a factor of 3-4 .

USE/ADVANTAGE - In microbiology, for producing anthrax vaccines. Savings in terms of reduced time and labour costs at spore material prodn. stage.

ABSTRACTED-PUB-NO: SU 1791449A
EQUIVALENT-ABSTRACTS:

CHOSEN-DRAWING: Dwg.0/0

DERWENT-CLASS: B04 C06 D16
CPI-CODES: B04-F10B; C04-F10B; B14-S11B; C14-S11B; D05-H07;

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End of Result Set

L1: Entry 10 of 10 File: DWPI Jan 23, 1993

DERWENT-ACC-NO: 1994-098617
DERWENT-WEEK: 199412

TITLE: Determining immunogenic activity of cattle anthrax vaccine - by feeding cattle pathogen spore suspension capsules in bread, and comparing test and control group survival

INVENTOR: AKHMEROV D SH,; NIZAMOV, R N ; SADYKOV, N S

PATENT-ASSIGNEE: VETERINARY RES INST (VETER)

PRIORITY-DATA: 1990SU-4806467 (February 7, 1990)

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PATENT-FAMILY:

	PUB-NO	PUB-DATE	LANGUAGE	PAGES	MAIN-IPC
☐	SU 1789218 A1	January 23, 1993		005	A61K039/00

APPLICATION-DATA:

PUB-NO	APPL-DATE	APPL-NO	DESCRIPTOR
SU 1789218A1	February 7, 1990	1990SU-4806467	

INT-CL (IPC): A61K 39/00

ABSTRACTED-PUB-NO: SU 1789218A
BASIC-ABSTRACT:

Determn. involves feeding cattle pathogen spore suspension capsules in bread in 0.75-0.90 x 10 power(9) spore dosage, then comparing test and control gp. survival.

A 100% mortality rate among non-vaccinated control gp. animals is guaranteed with the technique, which enables a rational approach to anti-anthrax immunisation to be developed. On the basis of controlled infection at various times after immunisation (15-30 days, 2-12 months) it was found possible to halve the frequency of vaccination (to a single procedure), thus achieving economies in vaccine prodn. costs.

USE/ADVANTAGE - Used in veterinary science, for evaluating prophylactic preparations, specifically anthrax vaccines for cattle. More accurate determn. of anthrax vaccine immunogenicity, leading to reduced frequency of immunisation.

In an example, after test animals had been immunised with the specific anthrax vaccine, both test and control gps. were infected by feeding them virulent anthrax pathogen suspensions in dosage of (0.75 x 10 power(9)) - (0.90 x 10 power(9))

spores at various times after immunisation (15, 21, 30 days, 2, 3, 6, 9, 12 months). months). The suspension is encapsulated in fast-dissolving gelatin and inserted in (5 x 5) - (10 x 10) cm pieces of bread. Vaccine effectiveness is assessed from the percentage of protected head.

ABSTRACTED-PUB-NO: SU 1789218A
EQUIVALENT-ABSTRACTS:

CHOSEN-DRAWING: Dwg.0/0

DERWENT-CLASS: B04 C06 D16
CPI-CODES: B04-F10B1; B11-C08E; B12-K04A4; B14-S11B; B14-S12; C04-F10B1; C11-C08E; C12-K04A4; C14-S11B; C14-S12; D05-H07; D05-H09;

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L5: Entry 1 of 24 File: PGPB Jun 17, 2004

DOCUMENT-IDENTIFIER: US 20040116370 A1
TITLE: Retroductal salivary gland genetic vaccination

Brief Description of Drawings Paragraph:
[0053] FIG. 12 illustrates a comparison of anti-anthrax protective antigen (PA) plasma IgG titers from retroductal introduction of formulations with DNA encoding PA with or without a polyionic organic acid into the salivary gland of rats.

Brief Description of Drawings Paragraph:
[0054] FIG. 13 illustrates a time course comparing anti-anthrax protective antigen (PA) plasma IgG titers using different introduction methods and positive (PA protein) and negative (hGH DNA) controls. Antibody titers were measured following retroductal delivery of PA DNA to the salivary gland (SG/PA DNA), injection of PA DNA into the muscle (i.m./PA DNA), or retroductal delivery of hGH DNA to the salivary gland (SG/hGH DNA). Subcutaneous PA protein plus CFA vaccination (s.c./Prtn+CFA), and naive animals served as positive and negative controls respectively. Arrows indicate when DNA or protein was administered.

Detail Description Paragraph:
Anti-Anthrax Protective Antigen Response Following Genetic Immunization

First Hit

L5: Entry 1 of 24 File: PGPB Jun 17, 2004

DOCUMENT-IDENTIFIER: US 20040116370 A1
TITLE: Retroductal salivary gland genetic vaccination

Brief Description of Drawings Paragraph:
[0053] FIG. 12 illustrates a comparison of anti-anthrax protective antigen (PA) plasma IgG titers from retroductal introduction of formulations with DNA encoding PA with or without a polyionic organic acid into the salivary gland of rats.

Brief Description of Drawings Paragraph:
[0054] FIG. 13 illustrates a time course comparing anti-anthrax protective antigen (PA) plasma IgG titers using different introduction methods and positive (PA protein) and negative (hGH DNA) controls. Antibody titers were measured following retroductal delivery of PA DNA to the salivary gland (SG/PA DNA), injection of PA DNA into the muscle (i.m./PA DNA), or retroductal delivery of hGH DNA to the salivary gland (SG/hGH DNA). Subcutaneous PA protein plus CFA vaccination (s.c./Prtn+CFA), and naive animals served as positive and negative controls respectively. Arrows indicate when DNA or protein was administered.

Detail Description Paragraph:
Anti-Anthrax Protective Antigen Response Following Genetic Immunization

First Hit

L5: Entry 7 of 24 File: PGPB Jan 15, 2004

DOCUMENT-IDENTIFIER: US 20040009945 A1
TITLE: Anthrax vaccine

Detail Description Paragraph:
[0033] The recombinant or fusion protein can be used as a vaccine for immunity against anthrax infection or as a diagnostic tool for detection of bacillus anthracis. The transformed host cells can be used to analyze the effectiveness of drugs and agents which inhibit anthrax or B. anthracis proteins, such as host proteins or chemically derived agents or other proteins which may interact with B. anthracis proteins of the present invention to inhibit its function. A method for testing the effectiveness of an anti-anthrax drug or agent can for example be the rat anthrax toxin assay (Ivins et al. 1984, Infec. Immun. 52, 454-458 and Ivins et al. 1986) or a skin test in rabbits for assaying antiserum against anthrax toxin (Belton and Henderson, 1956, Br. J. Exp. Path. 37, 156-160).

First Hit Fwd Refs

L5: Entry 18 of 24 File: USPT Jan 7, 2003

DOCUMENT-IDENTIFIER: US 6503906 B1
TITLE: Method for optimizing ciprofloxacin treatment of anthrax-exposed patients according to the patient's characteristics

Detailed Description Text (15):
It appears that the survival rate of the patients in the Sverdlovsk outbreak was similar to or slightly lower than the survival rate of the monkeys in the controlled group of the 1993 experiment. The Sverdlovsk patients were reportedly treated with penicillin, cephalosporin, chloramphenicol, anti-anthrax globin, corticosteroids, osmo-regulatory solutions, and artificial respiration. However, the exact dose regimens were not clearly described in the original paper [Meselson et al, 1994]. Out of the 68 deaths in the Sverdlovsk outbreak, 21 had documented onset-to-treatment time (FIG. 2), which did not show any significant treatment delay compared to the US data. The overall mortality rate of the Sverdlovsk patients (86%) was also similar to the occupationally acquired cases in the US (89%) [Inglesby et al, 1999], the later mostly occurring before the advent of antibiotics. Based on the fact that survival rate in the Sverdlovsk outbreak was similar to those of the animal controlled group and to the occupationally acquired cases without antibiotic treatment, it is apparent that the antibiotic treatment given to the Sverdlovsk patients was ineffective. Thus, the survival curve from the Sverdlovsk patients was treated as one obtained from an inactive controlled group.

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L5: Entry 19 of 24 File: USPT Aug 20, 2002

DOCUMENT-IDENTIFIER: US 6436933 B1
TITLE: Inhibitors of anthrax lethal factor activity

Brief Summary Text (11):
The pathogenesis of inhalational anthrax is more fully studied and understood. Inhaled spores are ingested by pulmonary macrophages and carried to hilar and mediastinal lymph nodes, where they germinate and multiply, elaborating toxins and overwhelming the clearance ability of the regional nodes. Bacteremia occurs, and death soon follows. Penicillin remains the drug of choice for treatment of susceptible strains of anthrax, with ciprofloxacin and doxycycline employed as suitable alternatives. Some data in experimental models of infection suggest that the addition of streptomycin to penicillin may also be helpful. Penicillin resistance remains extremely rare in naturally occurring strains; however, the possibility of resistance should be suspected in a biological warfare attack. Cutaneous anthrax may be treated orally, while gastrointestinal or inhalational disease ordinarily should receive high doses of intravenous antibiotics (penicillin G, 4 million units every 4 hours; ciprofloxacin, 400 mg every 12 hours; or doxycycline hyclate, 100 mg every 12 hours). The more severe forms require intensive supportive care and have a high mortality rate despite optimal therapy. The use of anti-anthrax serum, while no longer available for human use except in the the former Soviet Union, was thought to be of some use in the preantibiotic era, although no controlled studies were performed.

First Hit

L5: Entry 21 of 24 File: USOC Sep 28, 1965

DOCUMENT-IDENTIFIER: US 3208909 A
TITLE: Anaerobic process for production of a gel-adsorbed anthrax immunizing antigen antigen

OCR Scanned Text (3):
5 antigen and they were subseqtiently carefully examined in follow-up studies for any evidence of unusual or untoward clinical reactions to the antigen. No systemic reactions such as fever, headache, nausea, or anorexia were observed in any of the subjects. About 4% of the subjects exhibited mild local reactions at the site of the injection. These were primarily small localized areas of erythema and edema sometimes accompanied by slight itching. The reactions were mild and of a transitory nature, disappearing within 48 hours. The reaction tended to decrease after subsequent injections of the antigen in the three- injection initial immunization phase. Booster injections were given the volunteers six months later and eitber did not elicit any reactions or only mild reactions in those subjects that had previously shown a reaction during the initial immunization phase. These studies have shown unequivocally that the antigen is completely safe and is clinically acceptable for use in human immunization against anthrax. Studies were also made on the development of antianthrax antibody in the serum of the volunteer subjects immunized with the antigen. After the initial immunization series, 59% of the volunteers had serum antibody titers of 1-8 or higher. After the booster injection, 68% of the subjects had serum antibody titers of 1-8 Or higher. These studies show accordingly that the antigen is compatible with humans and is capable of developin.- a satisfacto.ry level of immunity, as indicated by serum antibody titers. It is to be understood that the specific medium used is one of preference only and that large variants are possible. Likewise variants are possible in the culturing, adsorption and stabilizing of the antigen. In considering the several alternatives available in medium composition it is to be noted that two factors generally are requi@red namely, organism growth and antigen elaboration into the medium. In addition, the rate of growth and therefore the time required for preparing and harvesting a batch of antigen becomes of importance. Growth is estimated by visual or other methods of observation of turbidity, and the elaboration of antigen is tested by assay of the lyophilized filtrate in guinea pigs by a standard in vivo assay. In addition, complement fixation titration is used as a standard in vitro assay to determine antigenic potency of culture filtrates. A pilot plant proving study has shown that the method as described herein is applicable for large scale production. Satisfactory potency was obtained in test lots of 300 liter volumes. It has been determined that sodium bicarbonate, calcium ion, leucine, isoleucine, proline, phenylalanine, methionine and histidine wore essential factors in antigen production. Essential also was a readily utilizable carbohydrate, such as glucose or sucrose. Similarly, magnesium ion was essential to growth and elements such as purines and vitamins were useful in stimulating and promoting growth. The presence of bicarbonate ' ion was shown to be necessary for the production of antigen and although the organism grew at a nornial rate when bicarbonate was omitted, or when the bicarbonate-free medium was aerated with carbon dioxide-free air, no detectable antigen was elaborated. When bicarbonate was omitted, the cultures became more acid than did control cultures during growth. When such cultures grown in bicarbonate-ftee medium were maintained at the same pH as the bicarbonate- containing control by periodic additions of ste-rile NAOH there was still no protective antigen in the cultures, or only traces of antigen were detected. Correspondingly, reduction of bicarbonate concentration

produced a decrease in antigen elaboration. Similarly, attempts to replace bicarbonate with compounds such as citrate, succinate, fumarate or maleate which a- re known 3,208,909 6 to be involved in carbon dioxide metabolism yesulted in no significant stimulation of antigen elaboration. Moreover, the addition of sodium citrate to a bicarbonate containing medium caused a marked inhibition of antigen elaboration, perhaps because of complex formation of metal ions in the medium. Thus, Thus, the need for bicarbonate ion for antigen elaboration appears to be conclusive. conclusive. It is significant also that the concentration of bicarbonate ion necessary for elaboration of antigen is relatively 10 high as compared with other components of the medium. It is of interest that isoleucine and phenylalanine are also necessary for productign of capsular polypeptide by Bacillus anthracis. Omission of glutamic acid delays but does not reduce the accumulation of polypeptide polypeptide and 15 antigen. Purines or their derivatives would not appear to be directly associated with elaboration of antigen, butrather to affect the process through stimulation of growth of t-he organism. Thus elaboration of antigen was detected in 20 the absence of purines if the incubation time was lengthened until good growth was obtained and the ability of the various purine compounds to enhance production of antigen was apparently correlated with their ability to stimulate growth. Threonine and glutamic acid were 25 apparently in a similar category. Omission of calcium chloride from the medium did not affect growth significantly, but catised a marked reduction in elaboration of protective antigen. Omission of magnesium sulfate resulted in almost complete inhibition 30 of growth. Doubling the concentrations of calcium and magnesium salts did not increase the elaboration of antigen. The organisms used @as production strains were obtained from certain wild-type strains isolated during na- 35 tural outbreaks of anthrax. Thus the three most satisfactory production strains used were secured from three of the parent strains isolated in various outbreaks in the United States. The wild-type strains were subjected to mtitation by exposure to ultra violet Tadiation, after 40 which t-he organisms were plated out and colonies selected that were rough, non- encapsulated, non-proteolytic and avirulent on animal inoculation. These strains were grown in the the preferred medium for a period of 40 to 45 hours at 37' C. The 45 hour maximum proved to be 45 the time after which degradation of the accumulated antiaen began to to set in. Prolonged incubation causes large antigen loss in the cultures, Samples of these three strains have been deposited with the ATCC. These have been given the designations ATCC 14185, 14186 50 and 14187. The preferred elements of the process rnay be enumerated as follows: (1) The growth of the culture is performed under anaerobic conditions in a chemically-defined, non-protein 55 medium. This method maintains the cultiire free of contaniination and haTMfUl foreign protein or other noxious substances and results in elaboration of a highpotency irnmlinizing antigen free from toxic material or contaminating viruses. 60 (2) Adsorption of the antigen onto a pre-formed aluminum hydroxide gel under specific conditions. This results in concentration of the antigenic material @and an increase in potency owing to the adjuvent effect of the 6,5 aluminum hydroxide gel. (3) Addition of Benzethonium Chloride Solution (XVI, 1960) and Formaldehyde Solution (XVI, 1960). These mate- rials give a preservative and a stabilizing ef- fect, respectively, to the final product so that freedom 7o from contamination and a high level of antigenic potency are maintained for extended periods of time. It is to be noted that the amount of formaldehyde solution added is far below that necessary to destroy or inactivate organisms, hence the flltrate or concentrate has to be sterile 75 prior to the above above additions.

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L11: Entry 3 of 138 File: PGPB Apr 22, 2004

DOCUMENT-IDENTIFIER: US 20040076638 A1
TITLE: Methods for preparing bacillus anthracis protective antigen for use in vaccines

CLAIMS:

20. A method for inducing serum antibodies which have neutralizing activity for B. anthracis toxin comprising administering to a mammal a pharmaceutical composition of of claim 19 comprising an amount of B. anthracis protective antigen sufficient to elicit production of said antibodies.

21. The method of claim 20 wherein the antibodies protect the human against infection by B. anthracis.

28. A method for inducing serum antibodies which have neutralizing activity for B. anthracis toxin comprising administering to a mammal a pharmaceutical composition of of claim 27 capable of producing an amount of B. anthracis protective antigen sufficient to elicit production of said antibodies in vivo.

29. The method of claim 28 wherein the antibodies protect the mammal against infection by B. anthracis.

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